## BME 695 Engineering Nanomedical Systems November 3, 2011 Copyright, 2011 – James F. Leary

## Lecture 14: Designing and Testing Integrated Nanomedical Systems

- 14.1 Introduction to integrated designs
  - 14.1.1 "Total design" but there is some order in the design process
  - 14.1.2 A brief outline of the total design process
- 14.2 Choose autonomous or non-autonomous design
  - 14.2.1 If autonomous, will there be error-checking to correct mistargeting?
  - 14.2.2 If autonomous, does the NMS perform all of the multi-step process sufficiently to accomplish the objective?
  - 14.2.3 If non-autonomous, what form of external modulation of the in-vivo nanomedical system will be used?
  - 14.2.4 If non-autonomous, are the external interactions able to adequately control the actions of the NMS?
  - 14.2.5 Evaluate reaction of NMS to external intervention
  - 14.2.6 Compare actions of NMS with and without external intervention.
  - 14.2.7 How do the actions of the NMS scale (linear? nonlinear? resonance? ) with the size or extent of the external intervention?
- 14.3 Choose core material, size and shape
  - 14.3.1 How will the core be used for diagnosis? Therapeutics?
  - 14.3.2 Does this dictate the core material? Size?
  - 14.3.3 Does shape alter circulation time or target cell penetration?
  - 14.3.4. Evaluate size and shape of nanosized core by transmission (TEM) or scanning electron microscopy (SEM), or by atomic force microscopy (AFM)
  - 14.3.5 Evaluate size of complete NMS (other parts may not be electron dense) by dynamic light scattering (DLS)
  - 14.3.6 Evaluate materials present at each layer of construction by x-ray photoelectron spectroscopy (XPS)
- 14.4 Design NMS targeting and evaluate its effectiveness
  - 14.4.1 Choose cell surface biomarker on diseased cell. Is it unique or just elevated in expression (e.g. folate receptors)
  - 14.4.2 Choose targeting molecule type (antibody, peptide, aptamer...)
  - 14.4.3 Use flow or image cytometry to evaluate correctness of targeting to diseased cell using that biomarker system
  - 14.4.4 How much mis-targeting is anticipated?What are the consequences of mistargeting?
  - 14.4.5 Determine degree of mistargeting and consider the costs of misclassification
  - (e.g. how many normal cells are mis-targeted for each diseased cell successfully targeted)
  - 14.4.6 Based on the costs of misclassification, reconsider additional or alternative diseased cell biomarkers?
  - 14.4.7 Evaluate intracellular targeting by TEM if NMS is not fluorescent)
  - 14.4.8 Evaluate intracellular targeting by 3D confocal fluorescence microscopy (if NMS is fluorescent)
  - 14.4.9 Evaluate intracellular targeting by 2D fluorescence microscopy if confocal microcopy is unavailable

- 14.5 Choose zeta potential
  - 14.5.1 Determine required zeta potential for outer/inner layers
  - 14.5.2 Determine pH of encountered microenvironments
  - 14.5.3 Determine ionic strength of encountered microenvironments
  - 14.5.4 Evaluate suitability of zeta potential
  - 14.5.5 If signs of agglomeration, modify zeta potential of NMS.
  - 14.5.6 Are the NMS sticking to any surfaces or cell types?
  - 14.5.7 Are the NMS being rapidly filtered by the kidneys in-vivo?
- 14.6 Choose stealth molecule
  - 14.6.1 Determine required time of circulation
  - 14.6.2 Circulation time will determine dose needed
  - 14.6.3 Evaluate effectiveness of stealth molecule
    - 14.6.3.1 Do the NMS show signs of protein deposition in-vitro or in-vivo?
    - 14.6.3.2 Are the circulation times of the NMS adequate to sufficiently target the diseased cells in-vivo?
- 14.7 Choose type and intracellular target of therapy
  - 14.7.1 Eliminate or fix the diseased cells?
  - 14.7.2 If choice is elimination, choose appropriate therapeutic molecule that will accomplish this action
  - 14.7.3 If choice is to fix the diseased cells, what therapeutic molecule can accomplish this action and how will it be controlled?
  - 14.7.4 Choose molecular measure of effectiveness of therapy (induced apoptosis, restoration of normal phenotype, ...)
  - 14.7.5 Use single cell analysis by flow cytometry to measure that molecular measure, if cells are in suspension.
  - 14.7.6 Use scanning image cytometry to measure that molecular measure, if cells are attached
- 14.8 A few final words on design of integrated nanomedical systems
  - 14.8.1 We are still in the early days of designing nanomedical systems. Some of the necessary feedback we need for better designs awaits early clinical trials on human patients and volunteers
  - 14.8.2 We do not understand some of the processes well enough to fully control their design. Still it is important to know what is important even if can not yet control it!

## References

1. Haglund, E., Seale-Goldsmith, M-M., Leary, J. F. "Design of Multifunctional Nanomedical Systems" Annals of Biomedical Engineering Annals of Biomedical Engineering, Vol. 37, No. 10, pp. 2048–2063 (2009).

2. Seale, M-M, Leary, J.F. "Nanobiosystems" WIREs (Wiley Interdisciplinary Reviews) Nanomed Nanobiotechnol 1: 553–567 (2009).